Pineapple

Robert E. Paull and Ching Cheng Chen Department of Plant and Soil Sciences, University of Hawaii at Manoa Honolulu, Hawaii

Scientific Name and Introduction: Pineapple (*Ananas comosus* L. Merr.) is a terrestrial member of the diverse family *Bromeliaceae*. The pineapple fruit has the distinction of being selected, developed and domesticated by peoples in tropical America in prehistoric times (Collins, 1968). Of the many pineapple cultivars, Smooth Cayenne is the major commercial cultivar. Other cultivars that are grown on a smaller scale include: Red Spanish, Queen, Pernambuco, Sugarloafs and Cabaiani. Low acid Smooth Cayenne varieties are also available (Nakasone and Paull, 1998).

Quality Characteristics and Criteria: Pineapple fruit must have a desirable size and shape, with flat "eyes" (individual fruitlets) and crown leaves that look fresh and are deep-green. High shell color is not always a good measure of sweetness. Negative characteristics include: dry, brown crown leaves; dull, yellow skin appearance; presence of mold on the surface or cut stem; and fruit having an unfirm feel.

Horticultural Maturity Indices: Pineapple fruit maturity is evaluated on the extent of fruit "eye" flatness and skin yellowing. Consumers similarly judge fruit quality by skin color and aroma. A minimum of 12% SSC is required for fresh fruit in Hawaii (Anon., 1968). A SSC:TA ratio of 0.9 to 1.3 is recommended (Soler, 1992a). Fruit do not continue to ripen or sweeten after harvest. Fully-ripe, yellow fruit are unsuitable for transporting to distant markets, so slightly less mature fruit are selected for this purpose (Akamine, 1963; Cancel, 1974). Immature fruit should not be shipped, since they do not develop good flavor, have low brix, and are more prone to chilling injury (Rohrbach and Paull, 1982).

Grades, Sizes and Packaging: Pineapples are graded by degree of skin coloration, size (weight), absence of defects and disease, and uniformity of these characteristics before packing. Other characteristics include maturity, firmness, nice shape, flat eyes, well-cured broken stem (peduncle), and a minimum SSC of 12% in Hawaii (Anon., 1968). Crown size is a crucial grade component, with a minimum size, and ratio of crown:fruit length (0.33 to 1.5) for higher grades. Crowns developed during the Summer in Hawaii tend to be larger and may require gouging (removal of the crown center) at harvest to meet the standard.

Pineapples are normally packed into cartons of two different sizes and on the basis of color and size: 1) a large telescoping fiberboard carton holding 18 kg (40 lb) and containing 8 to 10 fruit in two layers, flat or upright, for surface and air shipment; and 2) a smaller container of 9 kg (20 lb) with five to six fruit in a single layer laid flat is air shipped. Tourist packs of two to four fruit are also prepared. Absorbent pads are used at the bottom of the carton and between layers, if fruit are placed horizontally within the carton. In other packs, fruit are placed vertically.

Pre-cooling Conditions: Room-cooling or forced-air cooling should be used.

Optimum Storage Conditions: Temperatures of 7 to 12 °C (45 to 55 °F) are recommended for storage of pineapples for 14 to 20 days, provided fruit are at the color break stage (Paull, 1993). A RH of 85 to 95% is recommended; a high RH significantly reduces water loss. Ripe fruit can be held at 7.2 °C (45 °F) for about 7 to 10 days. Pineapples may be stored at 0 to 4 °C (32 to 39 °F) for weekss, but upon removal, fruit fail to continue ripening and show severe chilling injury. Quarter-yellow fruit at harvest gain about one additional week of storage for every 6 °C (11 °F) decrease in storage temperature (Dull, 1971). The maximum storage-life at 7 °C (45 °F) is about 4 weeks (Paull and Rohrbach, 1985). However, when

removed, chilling injury-induced internal browning develops within 2 to 3 days.

Controlled Atmospheres (CA) Consideration: Modified O_2 levels have shown only minimal effectiveness at extending pineapple shelf-life (Akamine and Goo, 1971; Dull et al., 1967). Some beneficial effect is gained from CA (4% O_2) treatments in reducing chilling injury development (Paull and Rohrbach, 1982). The fruit waxes currently used generate high internal concentrations of CO_2 (up to 5%) and reduced O_2 (Paull and Rohrbach, 1982). Low O_2 has no effect on crown condition or decay, but does delay shell color development and reduce superficial mold growth (Akamine and Goo, 1971). Subjecting fruit to 1 to 2% $O_2 + 0$ to 10% CO_2 after shipment from Mexico to England at 8 °C did delay chilling injury (Haruenkit and Thompson, 1994). Polyethylene bagging, though difficult to perform commercially on individual fruit, results in atmospheres of 8 to 10% $O_2 + 7$ % CO_2 , and fruit waxing delays appearance of chilling-induced internal browning (Abdullah et al., 1985; Paull and Rohrbach, 1982; Rohrbach and Paull, 1982). The tentative recommendation is 2 to 5% + 5 to 10% CO_2 (Yahia, 1998).

Retail Outlet Display Considerations: Whole fruit should be displayed refrigerated at 10 to 13 °C (50 to 55 °F). Do not mist or ice.

Chilling Sensitivity: Symptoms of chilling injury include: wilting; drying and discoloration of crown leaves; failure of green-shelled fruit to yellow; browning and dulling of yellow fruit; and internal flesh browning (Lim, 1985; Paull and Rohrbach, 1985). Pre-harvest shading, as well as pre-harvest and postharvest low temperature, are the major factors increasing symptom intensity (Akamine et al., 1975; Akamine, 1976; Keetch and Balldorf, 1979; Smith, 1983). Chilling injury symptoms include endogenous brown spot, physiological breakdown, blackheart, and internal browning, and they develop after fruit are returned to physiological temperatures of 15 to 30 °C (59 to 86 °F) (Paull and Rohrbach, 1985). Susceptible fruit are generally lower in ascorbate and sugar content and are opaque (Abdullah and Rohaya, 1983; Abdullah et al., 1985; Paull and Rohrbach, 1985; Swete Kelly and Bragshaw, 1993).

Ethylene Production and Sensitivity: The ethylene production rate of this non-climacteric fruit is low at 0.1 to 1.0 μL kg⁻¹ h⁻¹. Postharvest use of ethephon to degreen the shell has been tested in the Ivory Coast (Crochon et al., 1981; Poignant, 1971), Hawaii (Paull, 1985; unpublished data) and Australia (Smith, 1991). Treated fruit show more rapid uniform skin degreening and little change in quality. However, shelf-life is slightly shortened (Paull, 1985 unpublished; Smith 1991; Soler, 1992b). There is no recommendation for use of ethephon; it is not approved for postharvest use.

Respiration Rates:

Temperature	mg CO ₂ kg ⁻¹ h ⁻¹
5 °C	2
10 °C	4 to 7
15 °C	10 to 16
20 °C	19 to 29

To get mL kg⁻¹ h⁻¹, divide the mg kg⁻¹ h⁻¹ rate by 2.0 at 0 °C (32 °F), 1.9 at 10 °C (50 °F), and 1.8 at 20 °C (68 °F). To calculate heat production, multiply mg kg⁻¹ h⁻¹ by 220 to get BTU per ton per day or by 61 to get kcal per metric ton per day.

Physiological Disorders: Flesh translucency, also called porosity, is associated with greater fruit sensitivity to mechanical injury, indicated by leakage and oozing of cellular fluids. This condition begins before harvest and continues after harvest (Bowden, 1969; Rohrbach and Paull, 1982; Paull and Reyes, 1996). Fruit with increased translucency also have increased pH, a higher SSC/acid ratio, higher fruit weight, higher total esters, and lower acidity. SSC, flesh pigments, and palatability increase to a

maximum at about 60% translucency, then decline in fruit with higher translucency (Bowden, 1969). A related disorder is green shell ripe fruit having full translucency.

Bruising, due to impact damage, is a major problem during harvesting, packing and shipping of pineapple (Singleton, 1958). This injury is normally confined to the impact side of the fruit, and the damaged flesh appears slightly straw-colored (Keetch, 1978). Mechanical injury of translucent fruit can lead to leakage of cell contents and loss of marketable fruit. Injury can be avoided by careful handling and avoiding impacts and bruising.

Sunburn, or sun-scorched pineapples show a bleached, yellow-white skin that turns pale gray/brown with damage to the flesh underneath. Damaged areas are more susceptible to disease. It is common during hot periods, ie., > 35 °C (95 °F), of the year (Keetch and Balldorf, 1979).

Malformations, or pineapple fruit with pronounced eyes or fruitlets are normally not acceptable in Fancy grades of fruit, and the thicker skin results in lower flesh recovery. This condition is common in fruit that flower during cool weather. Some Spanish varieties are susceptible to broken core, in which the central core has a transverse break leading to the upper part of the fruit ripening ahead of the bottom (Lim, 1985).

Postharvest Pathology: Black rot, also called Thielaviopsis fruit rot, water blister, soft rot, or water rot, is a universal fresh pineapple problem characterized by a soft watery rot (Rohrbach, 1983). Diseased tissue turns dark in the later stages of the disease because of the dark mycelium and spores. Black rot is caused by the fungus *Chalara paradoxa* (De Seynes) Sacc. Red Spanish types are more resistant than 'Smooth Cayenne.' Infection occurs within 8 to 12 h following harvest and enters through the point of detachment or wounds. The severity of the problem is dependent on the degree of bruising or wounding during harvesting and packing, the level of inoculum on the fruit, and storage temperature during transportation and marketing (Rohrbach and Schmitt, 1994). The rot is commercially controlled by minimizing bruising of fruit during harvest and handling, refrigeration, and postharvest fungicides (Rohrbach and Phillips, 1990).

Fruitlet core rot, black spot, fruitlet brown rot, and eye rot describe the brown to black color of the central part of an individual fruitlet. Epidemic levels are rare in the major commercial pineapple-producing areas of the world (Rohrbach and Schmitt, 1994). Low-acid cultivars being grown commercially are most susceptible (Rohrbach and Schmitt, 1994). This disease is caused by a complex of fungi (Rohrbach and Schmitt, 1994). Infection frequently can lead to misshapen fruit that are culled before packing and shipping.

Yeasty fermentation, arises due to the fact that fruit are not sterile inside, containing many non-growing, but viable yeasts and bacteria. In damaged, overripe fruit and fruit with inter-fruitlet cracking, resident yeasts begin to grow, or new yeasts invade. This growth leads to fermentation and bubbles of gas and juice through cracks in the skin. The skin turns brown and leathery and fruit become spongy with bright yellow flesh.

Saprophytes growing on the broken end of peduncle (*Penicillium* sp.) and fruit surface are non-pathogenic but are unsightly, and therefore a marketing problem (Rohrbach, 1989). The condition is more common on highly translucent fruit.

Quarantine Issues: Pineapple fruit that are > 50% 'Smooth Cayenne' are not regarded a host for tephritid fruit flies. Thus, insect disinfestation is not required for import into fly-free countries (Armstrong, 1994).

Pineapple caterpillars (*Thecla basilides* Geyer; *Metamasius ritchiei* Marchall; *Batrachedre methesoni* Busch; *Paradiophorus crenatus* Billbarg) are exotic and limited to Central America, South America and the Caribbean (Harris, 1927; Rohrbach, 1983). The adult oviposits on the inflorescence prior to anthesis. Larvae then infest fleshy parts of the bracts and feed inside the developing inflorescence, exuding gum from the feeding chambers. Control with insecticides is relatively easy if flowering is induced uniformly with forcing agents. *T. basilides* is a tropical species that could cause

problems if imported into Southern states such as Florida, as it can feed on corn, cacao, Heliconia, and several other bromeliads (Rohrbach, 1983).

The pineapple scale, *Diaspis bromeliea* Kerner, occurs wherever pineapple is grown. Normally in Hawaii, pineapple scale is not a major problem in fields, probably because of scale parasites and predators. However, because of the quarantine requirement to have fruit insect free, even low levels of pineapple scale at harvest present quarantine problems. Scale are controlled by pre-harvest insecticide applications, taking into account "last-application-to-harvest" time.

The pineapple fruit mite, *Steneotarsonemus ananas* Tryon, occurs universally on the growing plant, developing inflorescence, fruit and crown. Fruit mites feed on developing trichomes on the white basal leaf tissue and flower bracts and sepals, causing light brown necrotic areas. The pineapple red mite, *Dolichotetranychus floridanus* Banks, feeds on the white basal leaf tissue, particularly of the crown. Severe damage occurs when the fruit mature under drought conditions. Red mites may cause death of basal crown leaves, affecting quality (Rohrbach and Schmitt, 1994).

Mealy bugs are removed from the surface by brushing. Pre-harvest insecticide and ant control almost eliminate mealy bugs (Soler, 1992a). Crickets/locusts may feed on bracts before harvest.

Suitability as Fresh-cut Product: Fresh-cut pineapple is readily prepared at a central plant or in-store as a ready-to-use consumer pack. Packs contain fresh-cut cylinders with the core removed, spears, chunks or wedges. The product has a shelf-life of at least 7 days at proper temperature. A patent has been issued for the use of pouches flushed with 15 to 20% O_2 + 3% argon and held at 1 °C (34 °F) for 10 weeks (Powrie et al., 1990), for fresh-cut pineapple pieces.

Special Considerations: Ease of removal of crown leaves, full skin yellowing or the sound produced by tapping the fruit are not signs of ripeness or quality. Fruit are picked at the ripe stage and are ready-to-eat, even if there is a little skin yellowing.

References:

- Abdullah, H. and M.A. Rohaya. 1983. The development of black heart disease in Mauritius pineapple (*Ananas comosus* cv. Mauritius) during storage at lower temperatures. MARDI Res. Bull. 11:309-319.
- Abdullah, H., M.A. Rohaya, and M.Z. Zaipun. 1985. Effect of modified atmosphere on black heart development and ascorbic acid contents in 'Mauritius' pineapple (*Ananas comosus* cv. 'Mauritius') during storage at low temperature. ASEAN Food J. 1:15-18.
- Akamine, E.K. 1963. Fresh pineapple storage. Hawaii Farm Sci. 12:1-4.
- Akamine, E.K. 1976. Postharvest control of endogenous brown spot in fresh Australian pineapples with heat. HortScience 11:586-588.
- Akamine, E.K. and T. Goo. 1971. Controlled atmosphere storage of fresh pineapples (*Ananas comosus* [L.] Merr. 'Smooth Cayenne'). Hawaii Agric. Exp. Stat. Res. Bull. No. 152, 8 pp.
- Akamine, E.K., T. Goo, T. Steepy, T. Greidanus and N. Iwaoka. 1975. Control of endogenous brown spot of pineapple in postharvest handling. J. Amer. Soc. Hort. Sci. 100:60-65.
- Anonymous, 1968. Wholesale standards for Hawaiian-grown pineapple, Hawaii Dept. Agric., Mkt. Div.
- Armstrong, J.W. 1994. Tropical and subtropical fruits. In: R.E. Paull and J.W. Armstrong (eds) Insect pests and fresh horticultural products: Treatments and responses. CAB Intl, Wallingford, U.K., pp. 275-290.
- Bowden, R.P. 1969. Further studies on ripeness in pineapple. Food Tech. Australia. 21:160-163.
- Cancel, H.L. 1974. Harvesting and storage conditions for pineapples of the 'Red Spanish' variety. J. Agric., Puerto Rico 58:162-169.
- Collins, J.L. 1968. The Pineapple. Leonard Hill, London. 295 pp.
- Crochon, M., R. Tisseau, C. Teisson and R. Huet. 1981. Effet d'une application d'Ethrel avant la recolte sur la qualite gustative de ananas de Cote d'Ivoire. Fruits 36:409-415.

- Dull, G.G. 1971. The pineapple: general. In: A.C. Hulme (ed) The biochemistry of fruits and their products. Vol. 2, Acad. Press, London, pp. 303-331.
- Dull, G.C., R. E. Young and J. B. Biale. 1967. Respiratory patterns in fruit of pineapple, *Ananas comosus*, detached at different stages of development. Physiol. Plant. 20:1059-1065.
- Harris, W.V. 1927. On a Lycaenid butterfly attacking pineapple in Trinidad, British West Indies. Bull. Ent. Res. 18:183-188.
- Haruenkit, R. and A.K. Thompson. 1994. Storage of fresh pineapple. In: B.R. Champ, E. Highley and G.I. Johnson (eds) Postharvest Handling of Tropical Fruits. Proc. Intl. Conf. Chiang Mai, Thailand, July 1993, ACIAR Proc No. 50, pp. 422-426.
- Keetch, D.P. 1978. Bruising of pineapples. Farming in South Africa Bulletin H11/1978, 2 pp.
- Keetch, D.P. and D. B. Balldorf. 1979. The incidence of certain pineapple fruit blemishes in the eastern cape and border. Citrus Subtrop. Fruit J. 551:12-15.
- Lim, W.H. 1985. Diseases and disorders of pineapples in Peninsular Malaysia. MARDI Rpt. No. 97, Kuala Lumpur, Malaysia, 53 pp.
- Nakasone, H.Y. and R.E. Paull. 1998. Tropical fruits. CAB Intl, Wallingford, U.K., 445 pp.
- Paull, R.E. 1993. Pineapple and papaya. In: G. Seymour, J. Taylor and G. Tucker (eds) Biochemistry of Fruit Ripening, Chapman & Hall, London, pp. 291-323.
- Paull, R.E. and K.G. Rohrbach. 1982. Juice characteristics and internal atmosphere of waxed 'Smooth Cayenne,' pineapple fruit. J. Amer. Soc. Hort. Sci. 107:448-452.
- Paull, R.E. and K.G. Rohrbach. 1985. Symptom development of chilling injury in pineapple fruit. J. Amer. Soc. Hort. Sci. 110:100-105.
- Paull, R.E. and M. E. Q. Reyes. 1996. Pre-harvest weather conditions and pineapple fruit translucency. Sci. Hort. 66:59-67.
- Poignant, A. 1971. La maturation controlee de l'ananas II-L'ethrel et son action au cours des phases ascendante et descendarte de la maturite. Fruits 26:23-35.
- Powrie, W.D., R. Chiu, H. Wu, and B.J. Skura. 1990. Preservation of cut and segmented fresh fruit pieces. U.S. Patent 4,895,729.
- Rohrbach, K.G. 1983. Exotic pineapple diseases and pests and their potential for spread. In: K.G. Singh (ed.) Exotic plant quarantine pests and procedures for introduction of plant materials. ASEAN Plant Quarantine Centre and Training Institute, Selangor, Malaysia, pp. 145-171.
- Rohrbach, K.G. 1989. Unusual tropical fruit diseases with extended latent periods. Plant Dis. 73:607-609.
- Rohrbach, K.G. and R. E. Paull. 1982. Incidence and severity of chilling induced browning of waxed 'Smooth Cayenne' pineapple. J. Amer. Soc. Hort. Sci. 107:453-457.
- Rohrbach, K. and D.J. Phillips. 1990. Postharvest diseases of pineapple. Acta Hort. 269:503-508.
- Rohrbach, K. and D.P. Schmitt. 1994. Pineapple. In: R.C. Ploetz, G.A. Zentmyer, W.T. Nishijima, K.G. Rohrbach and H.D. Ohr. (eds) Compendium of Tropical Fruit Diseases. Amer. Phytopath. Soc., St. Paul MN, pp 45-55.
- Singleton, V.L. 1958. A test for the degree of bruising of pineapple flesh. Pineapple Res. Inst. Hawaii, Honolulu. Pineapple Res. Inst. News 6:111-114 (Private document).
- Smith, L.G. 1983. Causes and development of black heart in pineapples. Trop. Agric. (Trinidad) 60:31-35.
- Smith, L.G. 1991. Effects of ethephon on ripening and quality of fresh market pineapples. Austr. J. Exp. Agric. 31:123-127.
- Soler, A. 1992a. Pineapple. CIRAD-IRFA, Paris, France, pp. 48, ISBN 2-87614-078-0.
- Soler, A. 1992b. Metabolism de l'ethephon dans l'epiderme de l'ananas (*Ananas comosus*, L. Merr). Fruits 47:471-477.
- Swete Kelly, D. and J. Bragshaw. 1993. Effect of fruit handling and fruit coatings on black heart (internal brown spot) and other aspects of fresh pineapple quality. Acta Hort. 334:305-316.
- Yahia, E. 1998. Modified and controlled atmospheres for tropical fruits. Hort. Rev. 22:123-183.